Amendments to the Specification:

Please amend the paragraph beginning at page 23, line 11 to page 24, line 2, as indicated in the following paragraph:

- - The RNA analysis was done according to Example 1 or the following procedure. Single stranded, biotinylated, DNA probes containing the specific HPV16 gene sequences were prepared. For HaCaT and SiHa cell lines, cells were grown to confluency, cells were harvested, and the total RNA was ~~isolated and~~ purified using the RNEASY® kit (Qiagen Inc., Santa Clarita, CA). For W12, whole cells were used for analysis. RNA calibrators containing the complete HPV genome were prepared by transcribing (+) sense RNA from a plasmid containing the complete HPV16 genome with T7 RNA polymerase. The RNA was then diluted to 10^3, 10^4, 10^5, 10^6, and 10^7 copies per 50 µl. Aliquots of cellular RNA were diluted to 50 µl and then 50 µl of Probe mix (containing the biotinylated, single-stranded DNA probe) was added to calibrators and cellular RNA and hybridized for 2 hours at 65 °C. The hybridization reactions were transferred to a streptavidin coated microplate and 25 µl of Detection Reagent 1 was added to each well. (Detection Reagent 1 contains the alkaline-phosphatase – anti-RNA:DNA monoclonal antibody conjugate.) During a 1 hour incubation with shaking, RNA:DNA hybrids were captured onto the streptavidin coated plate and were simultaneously reacted with the anti-hybrid antibody conjugate. After several wash steps, a chemiluminescent substrate (Tropix CDP-STAR® with Emerald) was added to the wells, and the light output was measured in a microplate luminometer after 30 minutes incubation at room temperature. - -